

Biological Control of Soilborne Pathogens (*Fusarium oxysporum* F. Sp. Cucumerinum) of Cucumber (*Cucumis sativus*) by *Trichoderma* sp.

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Abstract: In this study, the efficacy of the native isolates of *Trichoderma* species to promote the growth and yield parameters of cucumber and to manage *Fusarium* wilt disease under greenhouse conditions were investigated. Ninety native *Trichoderma* antagonists were isolated from Grassland and forest soil in different geographical regions of Inner Mongolia, China. Applications of *T. cf. harzianum* T1, T2, T3, T4, T5, T6, T7, T8, T10, T11 and *T. atroviride* (T9) exhibited the least disease incidence (by 0%) also the same strains shows 100% of relative control effect. Cucumber seedling treated with *T. cf. harzianum* (T2 and T1) isolates showed a significant stimulatory effect on plant height (by 13 and 14 cm respectively) and the highest shoot and root fresh weight were recorded by *T. asperelloides* (T27) and *T. gamsii* (T17) by 1.62 gm and 0.97 gm respectively, in comparison to untreated control and treated control (10 cm and 4 cm, 1.27 gm and 0.22 gm and 0.82 gm and 0.10 gm). Therefore, the antagonist (T1, T2, T3, T4, T5, T6, T7, T8, T9, T10, and T11) is chosen to be the most promising bio-control agent for *Fusarium oxysporum* f. sp. Cucumerinum and further study have to be exploited for sustainable disease management program.

Key words: Cucumber, biocontrol, *Fusarium*, *T. cf. harzianum*, *Trichoderma* spp.

1. Introduction

Cucumber (*Cucumis sativus* L.) is one of the most important economic crops, which belongs to family cucurbitaceae and is as important as a tomato in vegetable production in China. Cucumber is grown widely throughout the country, especially in North China, where it is the number one summer vegetable. China produces 70% of all the cucumbers in the world. According to data from the FAOSTAT, the statistical agency of the UN's FAO (Food and Agriculture Organization), with a total area of 1,150,000 ha and a total production of 48,000,000,000 kg. Cucumber is grown either in the open field or under protected houses. The purpose of growing crops under protected

house conditions is to extend their cropping season and to protect them from adverse conditions as well as diseases and pests [1].

Fusarium species are worldwide pathogenic fungi of many crop plants. Cucumber *Fusarium* wilt disease may occur at all growth periods of the cucumber plant [2]. The pathogens can survive as durable spores for many years with or without plant debris in soil, and it retains the ability to infect cucumber plants causing pre- or post-emergence damping-off, vascular discoloration of roots and stems, and eventually the entire plant wilts or dies. In China, an epidemic cucumber *Fusarium* wilt disease often occurred and led to a major yield loss in cucumber production. Generally, it caused cucumber yield losses of ~10% to 30% and poor quality products resulting in severe economic losses [3]. In China, cucumber *Fusarium* wilt was caused by several *Fusarium* species, including *F. oxysporum*, *F. equiseti*, *F. solani*, *F. moniliforme* and *F. proliferatum*. Among them, *F.*

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oxysporum is the major pathogen [4-6] and it is widespread in North China and is especially serious in glasshouse cropping systems [7]. *Fusarium oxysporum* f. sp. cucumerinum is an important soil-borne plant pathogen affecting cucumber plants (*Cucumis sativus* L.). Infected plantlets may develop root rot or stem canker, often leading to wilting and to plant death in severe cases [8].

Current cucumber disease management is based mainly on cultural practices and the use of fungicides, although chemical control is not always efficient or economically feasible for control of the disease. Biological control of the pathogen using mutualistic microorganisms has been suggested as a potential alternative to chemical control [9]. The biological control is the best alternative, especially against soil-borne pathogens. Biological control of pathogens, i.e., the total or partial destruction of pathogen populations by other organisms, occurs routinely in nature [10]. Among the various antagonists used for the management of plant diseases, *Trichoderma* spp. plays a vital role. These filamentous fungi are very common in nature, with high population densities in soil and plant litters [10]. Many studies have reported that *Trichoderma* spp. has the ability to reduce several plant diseases by inhibiting plant pathogens mainly found in the soil roots, through antagonistic and mycoparasitism potential [11]. For instance, studies conducted by Ref. [12] in Egypt and Ref. [13] in Vietnam, indicated that *Trichoderma* spp. had the ability to suppress the growth of fungal pathogens and enhance plant growth. However, no research has been conducted to evaluate the bio-efficacy of *T. cf. harzianum* in plant growth promotion and *Fusarium oxysporum* f. sp. cucumerinum disease management especially in cucumber seedling. Therefore, the objectives of this study were to evaluate the bio-efficacy of *Trichoderma* spp. with antagonistic activities against *Fusarium oxysporum* f. sp. cucumerinum and plant growth-promoting properties in cucumber seedling.

2. Materials and Methods

2.1 Isolation of *Trichoderma* spp.

Trichoderma spp. were isolated from the Grassland and forest soil of Inner Mongolia, China. Isolation of different *Trichoderma* spp. was made by the serial dilution technique. The soil solution was shaking for 30 min. And 1 ml of 10^{-3} dilution was poured onto *Trichoderma* selective Medium (MgSO₄: 0.20 g, KH₂PO₄: 0.90 g, NH₄NO₃: 1.0 g, KCl: 0.15 g, Glucose: 3.0 g, Rose Bengal: 0.15 g, Chloramphenicol: 0.25 g, streptomycin: 0.05 g, Agar: 15 g, Pentachloronitrobenzene (PCNB): 0.3 g, distilled water: 1 L) for selective isolation of *Trichoderma* and after the appearance of the colonies of *Trichoderma* on Petri dishes purified by hyphal tip isolation techniques. *Trichoderma* spp. was identified based on morphological and molecular methods using ITS5 (5'-GGAAGTAAAAGTCGTAACAAGG-3') and ITS4 (5'-TCCTCCGCTTATTGATATGC-3') according to Ref. [14] with modifications. The purified and identified cultures of *Trichoderma* spp. were maintained on PDA (Potato Dextrose Agar) medium and stored at 4 °C for further experimentation. Pure cultures were kept in 20% (w/v) glycerol at -20 °C.

2.2 Collection of Pathogens

Fusarium oxysporum f. sp. cucumerinum was obtained from China Academy of agricultural institute plant protection Research Laboratory (Beijing). The Isolate was maintained on PDA medium and placed until use at 4 °C.

2.3 Seed Treatment and Germination Testing

All tested cucumber seeds were disinfected using 10% sodium hypochlorite and soaked for 5 min then washed with distilled water for 4-5 times. After disinfection, the seeds are soaked in 3% (w/v) sodium carboxymethyl cellulose which serves as adhesive and stirring to improve the better stickiness of *Trichoderma*

spp., on the surface of the seed. The seed is coated with tested *Trichoderma* spp. and transferred to 90 mm petri dish with wet filter paper and each Petri plate will have 20 seeds then incubate at 28 °C for 5 days. Germinated seeds were recorded at the 5th day after incubation. Germination rate was calculated as the formula below:

Seed germination rate

$$= \frac{\text{No. of germinated seed}}{\text{Total No. of seeds}} \times 100$$

The experiment was conducted twice with five replicates per treatment. The germination rates were analyzed by Friedman test, LSD (Least Significant Different) [15].

2.4 Bio-Efficacy Evaluation of the Selected *Trichoderma* spp.

The bio-efficacy of the selected *Trichoderma* isolates was conducted under Glasshouse condition using a test tube. After the treated seed germinated 5 cucumber seedlings were transferred to 5 tubes

containing MS (Murashige and Skoog with vitamins) at a rate of 4.43 g/lit and kept for one day. The next day we added 2 mL of *Fusarium oxysporum* f. sp. cucumerinum and kept for 3 weeks. We use two different controls Ck1 the same treatment but without *Trichoderma* and without pathogen and Ck2 is with only *Fusarium oxysporum* f. sp. cucumerinum.

The disease severity, incidence and growth parameters of the cucumber seedlings in the test tube were evaluated at the vegetative stage: 3 weeks after transplanting (MacLean et al., 2002). The disease incidence was calculated by using the formula as described by Ref. [16]:

Disease incidence(%)

$$= \frac{\text{Total No. of infected plants per tube}}{\text{Total No. of plants per tube}} \times 100$$

The disease symptoms were evaluated based on to the disease scales from 0-5 (Table 1). The DSI (Disease Severity Index) was calculated following the calculation described by Ref. [17] using the following equation:

$$DSI(\%) = \frac{\sum(\text{Number on scale} \times \text{Number of seedlings in that scale})}{\sum(\text{Number of treated seedlings})(\text{highest rating value})} \times 100$$

$$\text{Reduction (\%)} = \frac{\text{Control} - \text{Treatment}}{\text{Control}} \times 100$$

Table 1 Cucumber seedling disease symptoms scoring scale.

Disease scale	Disease symptoms
0	Healthy and uninfected plants (no external symptoms)
1	Initial signs of wilting (yellowing)
2	Up to 25% of the leaves and root with symptoms
3	Up to 50% of the leaves and root with symptoms
4	Up to 75% of the leaves and root with symptoms
5	Plants dead

2.5 Statistical Analysis

All experiments were performed in five tubes per treatment and arranged in a randomized complete block design. Disease incidences (%) and disease severity index were analyzed using an ANOVA (analysis of variance) and grouped by Duncan test.

3. Result

3.1 Isolation of the Potential *Trichoderma* spp.

Total of 90 *Trichoderma* strains were isolated under 8 species from 76 soil samples collected from seven different regions of Xilingol, Hulunbuir, Tongliao, Chifeng, Ulanqab, Xing'an and Baotou within three Grassland and Forest Ecosystem in Inner Mongolia, Northern China. All isolated were confirmed as *Trichoderma* spp. based on morphological and Molecular results.

The isolated *Trichoderma* spp. is *T. cf. harzianum* (68), *T. longibranchiatum* (8), *T. asperelloides* (6), *T. afroharzianum* (2), *T. rossicum* (2), *T. hamatum* (1), *T. atroviride* (1) and *T. gamsii* (2).

To establish a phylogenetic tree, we first calculated haplotypes from 90 ITS5 and 4 sequences. Finally, 14

Table 2 Haplotypes (14) of *Trichoderma* identified in this study.

Haplotype	Species name	Number of strains	Representative strain	accession number
Hap1	<i>T.cf. harzianum</i>	12	T1	MF108890
Hap2	<i>T.cf. harzianum</i>	51	T5	MF108865
Hap3	<i>T. atroviride</i>	1	T9	-
Hap4	<i>T. gamsii</i>	2	T17	MF150096
Hap5	<i>T. asperelloides</i>	2	T63	MF114227
Hap6	<i>T. hamatum</i>	1	T34	MF144562
Hap7	<i>T. rossicum</i>	2	T23	MF144556
Hap8	<i>T. asperelloides</i>	4	T29	MF114228
Hap9	<i>T. longibrachiatum</i>	7	T64	MF102151
Hap10	<i>T.cf. harzianum</i>	1	T39	MF108907
Hap11	<i>T. afroharzianum</i>	2	T81	MF116243
Hap12	<i>T.cf. harzianum</i>	3	T57	MF108905
Hap13	<i>T. longibrachiatum</i>	1	T46	MF102164
Hap14	<i>T.cf. harzianum</i>	1	T50	MF109010

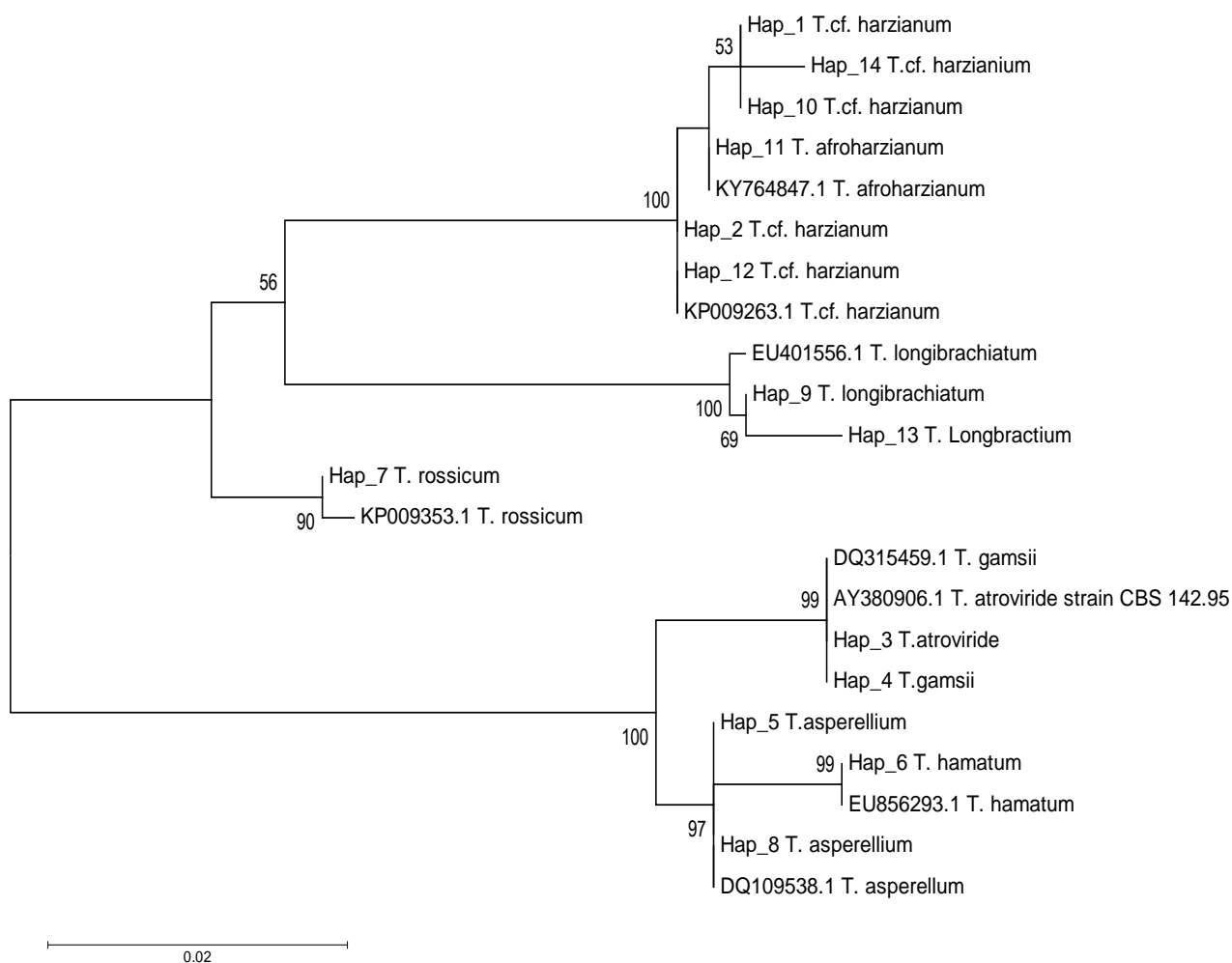


Fig. 1 Phylogeny reconstruction based on the ITS1-5.8S-ITS2 sequences from the rDNA (MEGA 6.0).

haplotypes (Table 2) were subjected to parsimony and Bayesian analysis. The result of this phylogenetic analysis is shown in Fig. 1. The 14 haplotypes belonging to nine *Trichoderma* species were positioned into seven clusters with strong bootstrap supports.

3.2 Effect of Treating Cucumber Seeds with *Trichoderma* Isolates on the Severity of *Fusarium* Wilt Disease

A total of ninety *Trichoderma* spp. strains were tested for their control efficiency against *Fusarium oxysporum* f. sp. *cucumerinum* on cucumber (*Cucumis sativus* L.). Data present in the Table 3 show that all tested isolates of *Trichoderma* spp. were antagonistic to *F. oxysporum* f. sp. *cucumerinum*. The disease incidence shows that the isolates have wide ranges from the lowest of 0 to the highest of 43. The relative control effect between tested strain was highly different compared to the control from the total tested strains. Eleven of them have reducing disease severity by 100% which consist two species which are ten strains of *T. cf. harzianum* and One strain of *T. atroviride* and the lowest was 53% which has been recorded by six strains of *Trichoderma* spp. under three species which are four strains of *T. cf. harzianum*, one strain of *T. rossicum* and one strain of *T. asperelloides*. In terms of growth parameters, significant differences were noticed between the tested isolates. The results on relative seed germination index on cucumber (*Cucumis sativus* L.) treated with *Trichoderma* isolates are presented in Table 3. The results show that the germination percentage ranges from the lowest of 85 to 96% indicating that the seeds were highly viable and the lower germination may be due to pre-emergence germination failure.

3.3 Screening the Relative Controlling Effect of the *Trichoderma* spp.

Eight *Trichoderma* spp., biological control agents i.e. *T. cf. harzianum*, *T. longibrachiatum*, *T. asperelloides*, *T. rossicum*, *T. afroharzianum*, *T. gamsii*, *T. atroviride* and *T. hamatum* were used to study their effect against

Fusarium oxysporum f. sp. *cucumerinum* i.e. (the most virulent isolates), the causal organism of cucumber wilt under greenhouse conditions. Data in Table 4 indicate that all tested biological control agents minimized the disease symptoms parameters in comparing with the infected control treatments. DI (disease incidence) was at lowest value by *T. atroviride* (0), followed by *T. hamatum* (7%) which both has one strain each while the highest DI value was recorded by *T. afroharzianum* (37%), *T. longibrachiatum* (28%) and *T. rossicum* (25%).

The highest number of strain isolated species was *T. cf. harzianum* (68) with average DI of 13%. The highest reducing disease severity was recorded by *T. atroviride* (100%) and *T. hamatum* (93%), followed by *T. cf. harzianum* (86%), while the lowest value was recorded by *T. afroharzianum* (60%), followed by *T. longibrachiatum* (70%). Table 4 indicates that all growth parameters of cucumber plants were improved by application of biological control agents against *Fusarium oxysporum* f. sp. *cucumerinum* compared with infested control treatments (Fig. 2). The highest shoot fresh weight was recorded by *T. asperelloides* (2 gm), followed by *T. hamatum* (1.30 gm) and *T. gamsii* (1.26 gm), while the lowest shoot fresh weight was recorded by *T. rossicum* (1.04 gm). Also, root fresh weight was at the highest value by *T. hamatum* (0.92 gm), followed by *T. gamsii* (0.87 gm), whereas the lowest root fresh weight was recorded by *T. rossicum* (0.56 gm). The biological control agents improved vegetative growth characters in treated cucumber plants. The highest vigor index was recorded by *T. atroviride* (936) followed by *T. hamatum* (902.5), while the lowest vigor index was recorded by *T. rossicum* (666.4).

3.4 Screening of the Relative Controlling Effect of *Trichoderma* Isolates Based on Their Isolation Regions and Ecosystem

The highest isolates were recorded in Typical Grassland Ecosystem with the rates of 71 strains

Table 3 Effect of Antagonistic fungi on disease incidence and growth parameters of FOC (F. O. f. sp. *cucumerinum*).

Trichoderma strains	Strain code	Disease incidence (%)	Relative control effect	Germination percent (%)	Growth parameters (gm fresh weight)		Vigor index
					Shoot	Root	
<i>T.cf. harzianum</i>	T1	0	100	96	1.31	0.85	1,324.8
<i>T.cf. harzianum</i>	T2	0	100	93	0.97	0.60	1,199.7
<i>T.cf. harzianum</i>	T3	0	100	93	1.12	0.66	1,106.7
<i>T.cf. harzianum</i>	T4	0	100	95	1.35	0.76	1,045.0
<i>T.cf. harzianum</i>	T5	0	100	96	1.18	0.75	1,048.3
<i>T.cf. harzianum</i>	T6	0	100	96	1.20	0.76	1,038.7
<i>T.cf. harzianum</i>	T7	0	100	95	1.18	0.82	1,018.4
<i>T.cf. harzianum</i>	T8	0	100	92	1.23	0.84	966.0
<i>T. atroviride</i>	T9	0	100	90	1.25	0.85	936.0
<i>T.cf. harzianum</i>	T10	0	100	92	1.45	0.91	947.6
<i>T.cf. harzianum</i>	T11	0	100	92	1.38	0.68	947.6
<i>T. longibrachiatum</i>	T12	3	96	92	1.03	0.78	938.4
<i>T.cf. harzianum</i>	T13	3	96	93	1.37	0.74	941.2
<i>T.cf. harzianum</i>	T14	3	96	93	1.28	0.78	937.4
<i>T.cf. harzianum</i>	T15	3	96	93	1.14	0.83	930.0
<i>T.cf. harzianum</i>	T16	3	96	89	1.13	0.75	882.9
<i>T. gamsii</i>	T17	3	96	93	1.43	0.97	920.7
<i>T.cf. harzianum</i>	T18	3	96	93	1.23	0.90	920.7
<i>T.cf. harzianum</i>	T19	3	96	92	1.21	0.69	910.8
<i>T.cf. harzianum</i>	T20	3	96	90	1.31	0.81	882.0
<i>T.cf. harzianum</i>	T21	7	93	93	1.52	0.68	911.4
<i>T.cf. harzianum</i>	T22	7	93	92	1.25	0.91	901.6
<i>T. rossicum</i>	T23	7	93	92	1.27	0.82	899.3
<i>T.cf. harzianum</i>	T24	7	93	90	1.13	0.81	878.4
<i>T.cf. harzianum</i>	T25	7	93	92	1.11	0.75	894.2
<i>T.cf. harzianum</i>	T26	7	93	95	1.16	0.84	921.5
<i>T. asperelloides</i>	T27	7	93	95	1.62	0.69	921.5
<i>T.cf. harzianum</i>	T28	7	93	95	1.21	0.77	919.6
<i>T. asperelloides</i>	T29	7	93	95	0.93	0.64	912.0
<i>T.cf. harzianum</i>	T30	7	93	92	1.33	0.75	877.7
<i>T.cf. harzianum</i>	T31	7	93	95	1.19	0.79	903.3
<i>T.cf. harzianum</i>	T32	7	93	95	1.27	0.77	902.5
<i>T.cf. harzianum</i>	T33	7	93	96	0.86	0.68	912.0
<i>T. hamatum</i>	T34	7	93	92	1.30	0.92	902.5
<i>T.cf. harzianum</i>	T35	7	93	92	1.05	0.87	874.0
<i>T.cf. harzianum</i>	T36	7	93	89	1.19	0.77	845.5
<i>T.cf. harzianum</i>	T37	10	89	92	1.35	0.72	872.2
<i>T.cf. harzianum</i>	T38	10	89	95	1.21	0.76	896.8
<i>T.cf. harzianum</i>	T39	10	89	92	1.32	0.70	866.6
<i>T.cf. harzianum</i>	T40	10	89	90	1.09	0.77	847.8
<i>T.cf. harzianum</i>	T41	10	89	90	0.88	0.59	846.0
<i>T.cf. harzianum</i>	T42	10	89	85	1.16	0.85	797.3
<i>T.cf. harzianum</i>	T43	10	89	95	1.05	0.73	891.1
<i>T.cf. harzianum</i>	T44	10	89	95	1.36	0.71	885.4
<i>T.cf. harzianum</i>	T45	10	89	95	1.03	0.84	883.5
<i>T. longibrachiatum</i>	T46	10	89	96	1.12	0.76	890.9

Table 3 continued

Trichoderma strains	Strain code	Disease incidence (%)	Relative control effect	Germination percent (%)	Growth parameters (gm fresh weight)		Vigor index
					Shoot	Root	
<i>T.cf. harzianum</i>	T47	10	89	92	1.33	0.70	846.4
<i>T.cf. harzianum</i>	T48	10	89	90	1.25	0.73	819.0
<i>T.cf. harzianum</i>	T49	10	89	95	1.34	0.92	864.5
<i>T.cf. harzianum</i>	T50	10	89	95	1.06	0.73	862.6
<i>T.cf. harzianum</i>	T51	10	89	89	1.32	0.78	805.5
<i>T.cf. harzianum</i>	T52	10	89	92	1.26	0.74	828.0
<i>T.cf. harzianum</i>	T53	10	89	89	0.66	0.84	801.0
<i>T.cf. harzianum</i>	T54	10	89	89	0.94	0.71	799.0
<i>T.cf. harzianum</i>	T55	10	89	92	0.97	0.85	818.8
<i>T.cf. harzianum</i>	T56	13	86	96	1.27	0.69	854.4
<i>T.cf. harzianum</i>	T57	13	86	92	1.25	0.76	816.5
<i>T.cf. harzianum</i>	T58	13	86	95	1.27	0.78	840.8
<i>T.cf. harzianum</i>	T59	13	86	95	1.39	0.79	836.9
<i>T.cf. harzianum</i>	T60	13	86	89	0.91	0.65	783.2
<i>T.cf. harzianum</i>	T61	13	86	92	1.06	0.94	809.6
<i>T.cf. harzianum</i>	T62	13	86	92	1.04	0.66	809.6
<i>T. asperelloides</i>	T63	17	82	90	1.16	0.75	790.2
<i>T. longibrachiatum</i>	T64	23	75	96	1.17	0.72	842.9
<i>T. asperelloides</i>	T65	23	75	85	1.21	0.80	744.8
<i>T. longibrachiatum</i>	T66	27	71	95	1.25	0.76	822.7
<i>T.cf. harzianum</i>	T67	27	71	95	1.26	0.76	817.0
<i>T. longibrachiatum</i>	T68	27	71	96	1.06	0.75	825.6
<i>T. longibrachiatum</i>	T69	27	71	95	1.32	0.77	807.5
<i>T. longibrachiatum</i>	T70	30	67	95	1.29	0.77	807.5
<i>T. longibrachiatum</i>	T71	30	67	95	1.03	0.77	807.5
<i>T. afroharzianum</i>	T72	33	64	95	1.13	0.75	807.5
<i>T. longibrachiatum</i>	T73	33	64	95	0.84	0.82	798.0
<i>T. longibrachiatum</i>	T74	33	64	95	1.03	0.61	788.5
<i>T.cf. harzianum</i>	T75	33	64	95	0.81	0.77	780.9
<i>T. asperelloides</i>	T76	33	64	95	1.29	0.79	779.0
<i>T.cf. harzianum</i>	T77	33	64	95	1.39	0.81	779.0
<i>T.cf. harzianum</i>	T78	33	64	85	1.23	0.80	688.5
<i>T.cf. harzianum</i>	T79	37	60	85	1.29	0.50	688.5
<i>T. longibrachiatum</i>	T80	37	60	95	1.09	0.73	765.7
<i>T. afroharzianum</i>	T81	40	57	95	1.35	0.74	750.5
<i>T.cf. harzianum</i>	T82	40	57	90	1.13	0.64	703.8
<i>T.cf. harzianum</i>	T83	40	57	93	1.09	0.81	716.1
<i>T.gamsii</i>	T84	40	57	95	1.09	0.77	731.5
<i>T.cf. harzianum</i>	T85	43	53	92	0.45	0.67	690.0
<i>T. asperelloides</i>	T86	43	53	93	0.98	0.59	585.9
<i>T.cf. harzianum</i>	T87	43	53	85	0.43	0.38	520.2
<i>T.cf. harzianum</i>	T88	43	53	85	0.80	0.67	493.0
<i>T.cf. harzianum</i>	T89	43	53	85	0.93	0.61	493.0
<i>T. rossicum</i>	T90	43	53	85	0.81	0.29	433.5
	Ck 1	0	100	96	1.3	0.8	938.4
	Ck 2	83	17	65	0.2	0.1	260.0

Table 4 Evaluation of eight *Trichoderma* spp., for the control of *Fusarium oxysporum* and growth performance on cucumber seedling under greenhouse conditions.

Species	No. of strains	DI (disease incidences)	Reducing disease severity (%)	Germination (G %)	Growth parameters (gm fresh weight)		Vigor index
					Root	Shoot	
<i>T. cf. harzianum</i>	68	13	86	92	0.75	1.15	863.43
<i>T. longibranchiatum</i>	8	28	70	95	0.74	1.10	815.5
<i>T. asperelloides</i>	6	22	76	92	0.78	2.00	788.9
<i>T. rossicum</i>	2	25	73	89	0.56	1.04	666.4
<i>T. afroharzianum</i>	2	37	60	95	0.75	1.24	779
<i>T. gamsii</i>	2	22	76	94	0.87	1.26	826
<i>T. atroviride</i>	1	0	100	90	0.85	1.25	936
<i>T. hamatum</i>	1	7	93	95	0.92	1.30	902.5
Control (Ck 1)		0	100	96	1.3	0.8	938.4
Control (Ck 2)		83	17	65	0.2	0.1	260.0
SEM		7.62	7.68	2.90	0.09	0.15	63.07

within 7 *Trichoderma* spp., i.e. *T. cf. harzianum* (52), *T. longibranchiatum* (8), *T. afroharzianum* (2), *T. asperelloides* (6), *T. hamatum* (1), *T. atroviride* (1), and *T. gamsii* (1) within seven regions of Inner Mongolia which are Chifeng (24), Xilingol (19), Ulanqab (10), Tongliao (7), Xing'an (6), Hulunbuir (3) and Baotou (2). The highest reducing disease severity was recorded by seven *T. cf. harzianum* (100%), five of them were isolated from Chifeng (T4, T5, T6, T7, and T10) and one each from Baotou (T8) and Xilingol (T11) and *T. atroviride* which were isolated from Hulunbuir (T9), while the lowest relative control was 52.9% recorded by 4 strains of *T. cf. harzianum* isolated from Xing'an (T85), Tongliao (T87), Chifeng (T88), Xilingol (T86) and *T. asperelloides* isolated from Tongliao (T86).

Meadow steppe has the second highest isolates rate with seven strains within four species *Trichoderma* spp., i.e. *T. cf. harzianum* (3), *T. rossicum* (2), *T. longibranchiatum* (1) and *T. gamsii* (1) from the regions of Xilingol, Chifeng and Xing'an. The highest reducing disease severity was recorded by *T. cf. harzianum* with rates of 100% with strains of T2 and T3 both are isolated from Xilingol. The lowest control effect was 52.9% with a strain of *T. rossicum* (T90) from Xing'an.

In Desert steppe all the strains were *T. cf. harzianum* isolated from 3 regions i.e. Chifeng,

Ulanqab, and Xilingol. The highest reducing disease severity was recorded by T1 at rates of 100% and the lowest control effect was recorded by four strains of T39, T40, T41, and T42 with the rates of 89%.

In forest, ecosystem 5 strains were isolated within two *Trichoderma* spp., from three regions of Xilingol, Hulunbuir and Xing'an. Three strains of *T. cf. harzianum* (T24, T38, and T47) and *T. longibranchiatum* (T68 and T69). The highest reducing disease severity was recorded by *T. cf. harzianum* (T24) at rates of 92.8%, while the lowest was recorded by two *T. longibranchiatum* (T68 and T69) with the rates of 71%.

3.5 Evaluating the Effectiveness of the Dominant *Trichoderma* spp., on *Fusarium Oxysporum* under in Vivo on Cucumber Seedling

The dominant *Trichoderma* spp. has shown significant antagonistic influence on the incidence of *Fusarium oxysporum* f. sp. cucumerinum on the seedling of cucumber. We have tested 68 strains of *T. cf. harzianum* (=H. 'pseudoharzianum' sensu Druzhinina et al., 2010a). We have found that from the total tested *T. cf. harzianum* strains 10 of them have 100% relative control effect with the strains of T1, T2, T3, ..., T8, T10, T11, which were isolated from three regions Chifeng (6), Xilingol (3) and Baotou (1) under two haplotypes of Hap1 (4 isolates) and Hap2 (6 isolates)



Fig 2 Effect of treating cucumber seeds with cell suspension antagonistic *Trichoderma* spp. isolates on the incidence of *Fusarium* wilt disease.

N.B. a-*T.cf. harzianum* (T1), b-Control 1 without any treatment and c-Control 2 inoculated with *F. oxysporum* f. sp. cucumerinum.

within three Ecosystem of Typical steppe (7), Meadow steppe (2) and Desert (1) while the lowest relative control effect was 53% recorded by 4 strains isolated from four regions xing'an, Tongliao, Chifeng, Xilingol and from single Ecosystem (Typical steppe) and Haplotype (Hap2).

The effect of various strains of *T. cf. harzianum* on the seed germination, vegetative growth, and vigor index of cucumber seedling was studied and the data are furnished in Table 2. Maximum germination percentage of cucumber seeds (96 percent,) was absorbed by six strains of T1, T5, T6, T33, T56, and T68, while the lowest Germination percentage rates were recorded by six strains of T42, T78, T79, T87, T88 and T89 at a rate of 85%. Regarding vegetative growth, the highest shoot and root weight were recorded by T21 (1.52 gm) and T61 (0.94 gm) respectively, while the lowest was recorded by T87 at a rate of the shoot (0.43 gm) and root (0.38 gm). The highest vigor index was shown by T1 (1324.3) and the

lowest was recorded by two strains of T88 and T89 at rates of 493.

4. Discussion

Trichoderma spp., bio controlling ability is related to its development of different mechanisms that allow it to compete directly for space and nutrients. In fungi, these mechanisms are associated with the production of antifungal metabolites, the secretion of hydrolytic enzymes, and their mycoparasitic ability, being behaviors that are expressed at different intensities depending on the strain used [18].

This study aimed to evaluate the efficacy of antagonistic *Trichoderma* isolates in controlling *Fusarium* wilt disease of cucumber under protected houses. Ninety (90) native strains under nine *Trichoderma* spp., were isolated from seven different regions under four ecosystems of Inner Mongolia soil and were characterized on the basis of their morphological and molecular features, effective in

reducing disease severity and the vegetative parameters were compared with the control and showed strong relative control effects which inhibited >53% infestation of *Fusarium oxysporum* f. sp. cucumerinum and eleven of the isolates showed 100% relative control of *Fusarium oxysporum* f. sp. cucumerinum while six isolates only showed 53% antagonistic potential.

All isolates were effective in reducing disease severity and increasing the average fresh weight of shoot and root. *T. cf. harzianum* (T1, T2, T3, T4, T5, T6, T7, T8, T10 and T11) has reduced disease severity by 100% apart from this *T. cf. harzianum* is a family of *Trichoderma harzianum* complex which was the dominant species isolated from different regions and ecosystem from the study area soil and the effective biological control in our trial. A similar trend was found by Ref. [19] when studying *T. harzianum* species, where it was concluded that each isolate could have a specific mycoparasitic activity. Therefore, it is not possible to generalize and indicate that a specific *Trichoderma* species can be classified as good antagonistic fungi because that characteristic is specific to each isolate. Sivakumar et al. [20] reported that the effects of *T. harzianum* were due to both antibiosis and mycoparasitism. Mechanisms employed by *Trichoderma* species to affect biological control of plant diseases are many and complex, and their use varies with the kind of biocontrol agent, pathogen, and host plant involved in the interaction [21]. Many investigators obtained similar results in greenhouse studies for controlling wilt disease on cucumber as well as other diseases [22-29].

Trichoderma atroviride is also well known as a biological control agent which reduces disease severity by (T9) 100%. This species has been confused with the closely related species *T. harzianum* because both species produce smooth, globose to sub-globose conidia. According to Ref. [30], the main difference between these two species is that *T. atroviride* produces a distinctive sweet or coconut

odor in culture, whereas *T. cf. harzianum* does not. We confirmed this specific odor from our isolated *T. atroviride* strains and did not detect it from any of the isolated *T. cf. harzianum* strains. On the other hand *T. cf. harzianum* (T85, T87, T88, and T89), *T. asperelloides* (T86) and *T. rossicum* (T90) were the least effective in reducing disease severity by 53%. It was reported that isolates of the *T. harzianum* collected from different soil samples were not equally effective to inhibit the growth of *F. udum* [31, 32]. It was found that one isolate (T1) among the 10 isolates of this antagonist was effective. So, there is need to search a very specific isolate(s) of *Trichoderma* sp. for successful control of *Fusarium oxysporum* f. sp. lycopersici [32].

As per result, ninety (90) potential isolates of *Trichoderma* may be further exploited as a biocontrol agent against *Fusarium oxysporum* f. sp. cucumerinum, as well as other soil, bore phytopathogenic fungi. In the present study, the better efficacy was observed in treatments including *T. cf. harzianum* (Table 3).

5. Conclusion

By using *Trichoderma* spp. such as *T. cf. harzianum*, *T. atroviride* and *T. gamsii* we could reduce disease incidence percentage of cucumber exposed to *Fusarium* pathogens such as *Fusarium oxysporum* f. sp. cucumerinum as agents of root and stem rot cucumber under greenhouse conditions. The presented method of bio controlling of pathogens can be considered as an applicable strategy in control measures against pathogens.

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Competing Interests

The authors declare that they have no competing interests.

References

- [1] Hanam, J. J., Holley, W. D., and Goldsberry, K. L. 1978. *Greenhouse Management*. Berlin: Springer-Verlag.
- [2] Celetti, M. 2007. *Fusarium* Wilts in Cucumber. <http://www.omafra.gov.on.ca/english/crops/hort/news/vegnews/2007/vg0907a4.htm>.
- [3] Li, J., Yang, Q., Zhang, S. M., Wang, Y. X., and Zhao, X. Y. 2009. "Evaluation of Biocontrol Efficiency and Security of a *Bacillus Subtilis* Strain B29 against Cucumber *Fusarium* Wilt in Field." *China Vegetables* 2: 30-3.
- [4] Huang, Z. X. 1990. "Occurrence of Cucumber Wilt Disease and Identification of Pathogen in Shanghai Suburbs." *Acta Agriculture Shanghai* 6 (2): 57-62.
- [5] Huang, Z. S., and Yang, Y. R. 1990. "Study on Cucumber *Fusarium* Wilt Pathogen and Its Control." *Acta Agriculture Boreali-Sinica* 5 (4): 99-104.
- [6] Chen, X., Liu, D., Zhang, Y. J., Qin, Z. W., and Zhou, X. Y. 2010. "Isolation and Identification of *Fusarium* from Cucumber Wilt Plants." *Journal of Northeast Agricultural University* 41 (7): 37-44.
- [7] Huang, Z. S., Yang, Y. R., and Zhu, X. D. 1994. "Identification of Pathogenic Races and Integrated Control of *Fusarium* Wilt of Cucumber in China." *Acta Agriculture Boreali Sinica* 9 (4): 81-6.
- [8] Rose, S., and Parker, M. 2003. "Efficacy of Biological and Chemical Treatments for Control of *Fusarium* Root and Stem Rot on Greenhouse Cucumber." *Plant Disease* 87: 1462-70.
- [9] Park, C. S., Paulitz, T. C., and Baker, R. 1988. "Biocontrol of *Fusarium* Wilt of Cucumber Resulting from Interactions between *Pseudomonas Putida* and Nonpathogenic Isolates of *Fusarium Oxysporum*." *Phytopathology* 78 (2): 190-4.
- [10] Akrami, M., and Yousefi, Z. 2015. "Biological Control of *Fusarium* Wilt of Tomato (*Solanum Lycopersicum*) by *Trichoderma* spp. as Antagonist Fungi." *Biological Forum—An International Journal* 7 (1): 887-92.
- [11] Viterbo, A., Landau, U., Kim, S., Chernin, L., and Chet, I. 2010. "Characterization of ACC Deaminase from the Biocontrol and Plant Growth-Promoting Agent *Trichoderma Asperellum* T203." *FEMS Microbial. Lett.* 305: 42-8.
- [12] Al-Tuwaijri, M. M. Y. 2015. "Studies on *Fusarium* Wilt Disease of Cucumber." *Journal of Applied Pharmaceutical Science* 5 (2): 110-9.
- [13] Ha, T. N. 2010. "Using *Trichoderma* Species for Biological Control of Plant Pathogens in Vietnam." *J. ISSAAS* 16: 17-21.
- [14] Samuels, G. J., Chaverri, P., Farr, D. F., and McCray, E. B. 2009. *Trichoderma* Online Systematic Mycology and Microbiology Laboratory Homepage at: <http://ntars-gringov/taxadescriptions/keys/TrichodermaIndexcfm>.
- [15] Bandyopadhyay, R., Cardwell, K. F., and Neuenschwander, P. 2003. "Species of *Trichoderma* and *Aspergillus* as Biological Control Agents against Plant Diseases in Africa." In *Biological Control in IPM Systems in Africa*, edited by Neuenschwander, P., Borgemeister, C., and Langewald, J. Wallingford: CABI Publisher. ISBN: 13-9781845933388, 193-206.
- [16] Teng, P. S., and James, W. C. 2001. "Disease and Yield Loss Assessment." In *Plant Pathologist's Pocketbook*, edited by Waller, J. M., Lenne, J. M., and Waller, S. J. Boston, Massachusetts: CABI Pub. Co. Inc., 25-38.
- [17] Ooi, K. H. 2002. "Chemical Characterization and Control of *Fusarium oxysporum*, Causes Vascular Wilt Disease in Rosel." Ph.D. Thesis, University Sains Malaysia, Malaysia.
- [18] Michel Aceves, A. C., Otero Sánchez, M. A., Ariza Flores, R., Barrios Ayala, A., and Alarcón Cruz, N. 2013. "Eficiencia biológica de cepas nativas de *Trichoderma* spp., en el control de *Sclerotium rolfsii* Sacc., en cacahuete." *Avances Investig Agropec* 17: 89-107.
- [19] Hoyos-Carvajal, L., Chaparro, P., Abramsky, M., Chet, L., and Orduz, S. 2008. "Evaluación de aislamientos de *Trichoderma* spp. contra *Rhizoctonia solani* y *Sclerotium rolfsii* bajo condiciones in vitro y de invernadero." *Agron Colombiana* 26: 451-8.
- [20] Sivakumar, D., Wilson Wijeratnam, R. S., Wijesundera, R. L., Marikar, F. M., and Abeyesekere, M. 2000. "Antagonistic Effect of *Trichoderma harzianum* on Postharvest Pathogens of Rambutan (*Nephelium lappaceum*)." *Phytoparasitica* 28 (3): 240-7.
- [21] Howell, C. R. 2003. "Mechanisms Employed by *Trichoderma* Species in the Biological Control of Plant Diseases, the History and Evolution of Current Concepts." *Plant Dis.* 87: 4-10.
- [22] Ercole, N., and Gennari, S. 1993. "Biological Control of *Fusarium* Wilt of Melon by Seed Coating with *Trichoderma Harzianum*." *Rifai. Culture Protecte* 22: 73-4.
- [23] Lifshitz, R., Windham, M. T., and Baker, R. 1986. "Mechanism of Biological Control of Pre-emergence Damping-off of Pea by Seed Treatment with *Trichoderma* spp." *Phytopathology* 76 (7): 720-5.

- [24] Simochkina, V. I., Uspanov, A. K., and Bekmakhanova, N. E. 1988. "Use of *Trichoderma* to Control Root Rot of Cucumber Undercover." *Izvestiya Akademii Nauk Kazakhskoi SSR Seriya Biologicheskaya* (5): 48-52.
- [25] Khalifa, E. Z. 1991. "Biological Control of Tomato *Fusarium* Wilt by *Trichoderma Harzianum*." *Menofiya. J. Agric. Res.* 16: 1246-59.
- [26] Lizuosen, H., Yue, Q., and Xia, X., 2004. "Inhibitory Spectrum and Partial Biological Traits of Five *Trichoderma* Isolates." *[Chinese] Journal of Yunnan Agricultural University* 19 (3): 267-71.
- [27] Zhu, T. H., Xing, X. P., and Sun, S. 2004. "The Antagonism Mechanisms and Diseases Control Trials of *Trichoderma* Strain T97 against Several Plant Fungal Pathogens in the Greenhouse." *Acta Phytophylacica Sinica* 31 (2): 139-44.
- [28] Rose, S., Yip, R., and Punja, Z. K. 2004. "Biological Control of *Fusarium* and Pythium Root Rots on Greenhouse Cucumbers Grown in Rock Wool." *Acta Horticulturae International Society for Horticultural Science (TSHS)* 635: 73-8.
- [29] Awad, H. M. 2004. "Studies on Root Rot Diseases of Pea." M.Sc. thesis, Faculty of Agriculture, Minufiya University.
- [30] Dodd, S. L., Lieckfeldt, E., and Samuels, G. J. 2003. "Hypocrea Atroviridis sp. nov., the Teleomorph of *Trichoderma Atroviride*." *Mycologia* 95: 27-40.
- [31] Bell, D. K., Well, H. D., and Markham, C. R. 1982. "In Vitro Antagonism of *Trichoderma* Species against Six Fungal Plant Pathogens." *Phytopathology* 72: 379-82.
- [32] Biswas, K. K., and Das, N. D. 1999. "Biological Control of Pigeon Pea Wilt Caused by *Fusarium* Udum with *Trichoderma* spp." *Ann. Plant Protection Science* 7 (1): 46-50.